We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

4,800
Open access books available

122,000
International authors and editors

135M
Downloads

Our authors are among the

154
Countries delivered to

TOP 1%
most cited scientists

12.2%
Contributors from top 500 universities

CLARIVATE ANALYTICS BOOK CITATION INDEX INDEXED

WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.
For more information visit www.intechopen.com

Chapter 2

Fundamentals of Medicinal Application of Titanium Dioxide Nanoparticles

Kazutaka Hirakawa

Additional information is available at the end of the chapter

http://dx.doi.org/10.5772/61302

Abstract

Titanium dioxide (TiO_2), a semiconducting material, is a well-known photocatalyst. A nanoparticle (NP) of TiO_2 also demonstrates photocatalytic activity. Photo-irradiated TiO_2 NPs induce the formation of various reactive species, leading to the damage of biomacromolecules. These reactive species include h^+, either free or trapped hydroxyl radicals ($OH^·$), superoxide ($O_2^{·-}$), hydrogen peroxide (H_2O_2), and singlet oxygen ($^1O_2$), among others. TiO_2 NPs photocatalyze DNA oxidation. A relatively small concentration of TiO_2 NPs frequently induces tandem base oxidation at guanine and thymine residues through H_2O_2 generation in the presence of a copper(II) ion. A copper–peroxo complex is considered to be an important reactive species responsible for this DNA damage. In the case of a high concentration of TiO_2 NPs, $OH^·$ contributes to DNA damage without sequence specificity. In the presence of sugars, TiO_2 NPs indirectly induce DNA damage by the secondary H_2O_2, which is produced through an autoxidation process of the product of sugar photooxidized by TiO_2 NPs. Furthermore, $^1O_2$ is also produced by photo-irradiated TiO_2 NPs. The photocatalyzed formation of $^1O_2$ might contribute to the oxidation of the membrane protein. These mechanisms of photocatalytic formation of the reactive species may be involved in the photocytotoxicity of TiO_2 NPs.

Keywords: Titanium dioxide, Photocatalyst, Reactive oxygen species, Photomedicine, DNA damage

1. Introduction

Titanium dioxide (TiO_2), a semiconducting material, is a well-known photocatalyst [1-5]. Examples of previous studies about TiO_2 photocatalytic reactions are listed in Table 1. A nanoparticle (NP) of TiO_2 also demonstrates photocatalytic activity. Important applications of TiO_2 photocatalysts are bactericidal activity [2-4, 6-12] and degradation of chemical pollutants

[2-4, 13]. Related physical and chemical mechanisms have been also investigated [2-5, 14-17]. Photo-irradiated TiO_2 NPs induce the formation of various reactive species, leading to the damage of biomacromolecules. These reactive species include hole (h^+), either free or trapped hydroxyl radicals ($OH^·$), superoxide ($O_2^{·-}$), hydrogen peroxide (H_2O_2), and singlet oxygen ($^1O_2$), among others. Hydroxyl radicals, $O_2^{·-}$, H_2O_2, and $^1O_2$ are the typical reactive oxygen species. TiO_2 photocatalysts have been found to kill cancer cells [18-21] other than bacteria, viruses, and algae under ultraviolet-A (wavelength: 315–400 nm) illumination [2-4, 6-12]. Therefore, one of the potential applications of the TiO_2 NP photocatalyst is photodynamic therapy (PDT), which is a promising treatment for cancer and some nonmalignant conditions [22-25]. In general, the mechanism of cytotoxicity by the photocatalysis of TiO_2 is based on cell membrane damage via the generation of the aforementioned reactive oxygen species. Furthermore, DNA damage in human cells [26-28], mouse lymphoma cells [29], and phage [30] by the TiO_2 NP photocatalyst has been reported. Direct damage of isolated DNA by TiO_2 photocatalyst *in vitro* has been also studied [31, 32]. However, the DNA-damaging mechanism *in vivo* is not well-understood, because the incorporation of the TiO_2 NPs in the nucleus is difficult [18]. A previous study has shown that H_2O_2 formation through the photocatalytic reaction of TiO_2 may contribute to cellular DNA damage [2, 19]. Hydrogen peroxide, a long-lived reactive oxygen species, can penetrate the nucleus membrane and induce oxidation of the nucleobase and strand breakage through enhancement by metal ions. Iron or copper ions can enhance the activity of H_2O_2 to produce $OH^·$ [33] and copper-peroxide [34-36]. Furthermore, secondary generation of reactive oxygen species may contribute to cytotoxicity of TiO_2 NPs photocatalyst [37]. Since the photocatalytic reaction will occur in a complex biological environment, an interaction between TiO_2 NPs and biomaterials should participate in the generation of reactive species to induce DNA damage. For example, sugars photocatalyzed by TiO_2 NPs may secondarily generate H_2O_2 through their further oxidation process by molecular oxygen in the presence of a metal ion [37]. In addition, the possibility of $^1O_2$-mediated cytotoxicity by TiO_2 NPs has been proposed [38]. Actually, $^1O_2$ generation by photo-irradiated TiO_2 NPs was demonstrated by a near-infrared spectroscopy [39, 40]. In this chapter, recent studies about photocatalytic biomacromolecule damage by TiO_2 NPs are briefly reviewed.

Target	References
Reviews	[2], [3], [4], [5]
Physical experiment	[1], [16], [17], [39], [40]
Chemical compounds	[13], [14], [15]
Nucleic acids	[31], [32]
Microorganism	[6], [7], [8], [9], [10], [11], [12], [30]
Cancer cell	[18], [19], [20], [21]
Mouse lymphoma cells	[29]
Cancer treatment of mouse	[20]

Table 1. Summary of the examples of previous studies onTiO_2 photocatalyst

http://dx.doi.org/10.5772/61302

1.1. General mechanism of photocatalysis of TiO_2 NP

The crystal of TiO_2 is a semiconductor, and the two crystalline forms, anatase and rutile, are well-known (Figure 1) [2-5]. The values of the band gap energy of these crystal forms are 3.26 and 3.06 eV for anatase and rutile, respectively. Photo-irradiation to a TiO_2 crystal induces the formation of an excited electron (e^-) in the conduction band and an h^+ in the valence band, leading to the redox reaction of materials adsorbing on the TiO_2 surface, including water and/or molecular oxygen. The photocatalytic reactions with its surface water and oxygen cause the formation of various reactive oxygen species such as free or trapped $OH^\cdot$, $O_2^{\cdot-}$, H_2O_2, and $^1O_2$ [2-5].

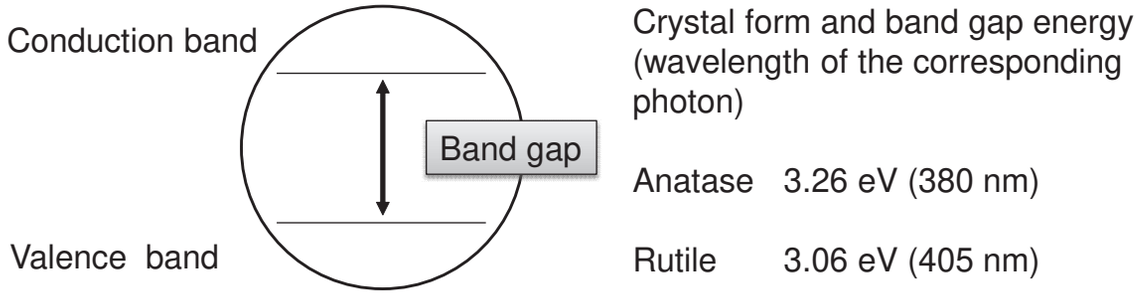

Figure 1. Band gap energy of the two crystalline forms of TiO_2.

An excited electron in the conductive band reduces the oxygen molecule adsorbed on the surface of TiO_2 NPs, leading to the generation of various reactive oxygen species as follows (Figure 2):

$$O_2 + e^- \rightarrow O_2^{\cdot-} \quad (1)$$

$$2O_2^{\cdot-} + 2H^+ \rightarrow H_2O_2 + O_2 \quad (2)$$

$$H_2O_2 \rightarrow 2OH^\cdot \quad (3)$$

The reaction (3) is mediated by ultraviolet radiation (hν, wavelength <355 nm), metal ions (M^{n+}) such as Fe^{2+}, and $O_2^{\cdot-}$, as follows [33]:

$$H_2O_2 + h\nu \rightarrow 2OH^\cdot \quad (4)$$

$$H_2O_2 + M^{n+} \rightarrow OH^\cdot + OH^- + M^{(n+1)+} \quad (5)$$

$$H_2O_2 + O_2^{\cdot-} \rightarrow OH^{\cdot} + OH^{-} + O_2 \quad (6)$$

On the other hand, the formed h^+ in the valence band can oxidize water to form $OH^{\cdot}$ as follows:

$$H_2O + h^+ \rightarrow OH^{\cdot} + H^+ \quad (7)$$

Furthermore, $OH^{\cdot}$ can produce H_2O_2 as follows:

$$2OH^{\cdot} \rightarrow H_2O_2 \quad (8)$$

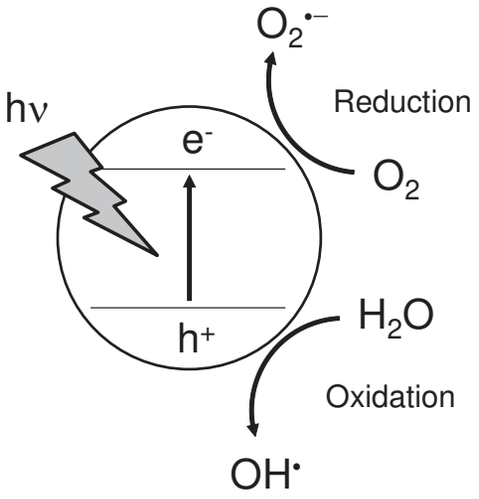

Figure 2. Photocatalytic reactive oxygen formation by TiO_2.

A photo-irradiated TiO_2 NP can induce $^1O_2$ formation. The formation of $^1O_2$ is considered to be an important mechanism of PDT. This reaction can be explained by the following process: $O_2^{\cdot-}$ formed by TiO_2 photocatalysis is reoxidized by the h^+ of TiO_2 on the particle surface to form $^1O_2$ as follows (Figure 3):

$$O_2^{\cdot-} + h^+ \rightarrow {}^1O_2\left({}^1\Sigma_g^{+} \text{ or } {}^1\Delta_g\right)\left(\text{or } {}^3O_2\right) \quad (9)$$

$$^1O_2\left({}^1\Sigma_g^{+}\right) \rightarrow {}^1O_2\left({}^1\Delta_g\right) \quad (10)$$

These reactive oxygen species should contribute to the mechanism of the phototoxicity induced by TiO_2 NPs.

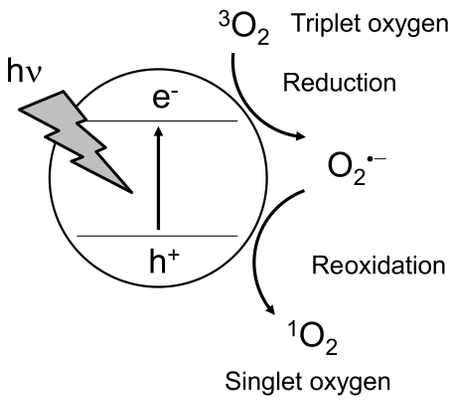

Figure 3. Photocatalytic $^1O_2$ generation by TiO_2

1.2. Sterilization effect by TiO_2

One of the most important medicinal applications of TiO_2 NPs is to kill bacteria on its surfaces. TiO_2 NPs under ultraviolet radiation produce a strong oxidative effect through the formation of above-mentioned reactive oxygen species and can be used as a photocatalytic disinfectant without other chemical reagents. Fujishima and coworkers reported the bactericidal effect of TiO_2 photocatalysts against *Escherichia coli* under ultraviolet-A irradiation using black light [6]. This is the first report of the application of phototoxicity of TiO_2 NPs. It was speculated that H_2O_2 was a reactive species responsible for this phototoxic effect [7]. Relevantly, the photocatalytic effect of TiO_2 against methicillin-resistant *Staphylococcus aureus* (MRSA) and *Clostridium difficile* in hospitals has been reported [10]. The bactericidal effect of TiO_2 NPs could be enhanced by metal doping [9]. Furthermore, visible-light-induced TiO_2 photocatalysts were developed and utilized in antibacterial applications. For example, sulfur-doped TiO_2 demonstrates the killing effect on *Escherichia coli* under white-light irradiation commonly used in hospitals [11].

1.3. Photodynamic therapy

Photodynamic therapy, which is a promising and less-invasive treatment for cancer, employs a photosensitizer and visible light to produce oxidative stress in cells and ablate cancerous tumors [22-25]. Photodynamic therapy is also used for treating some nonmalignant conditions that are generally characterized by the overgrowth of unwanted or abnormal cells. In general, porphyrins are used as photosensitizers under visible-light irradiation, since the human tissue has relatively high transparency for visible light, especially red light, and visible light has hardly any side effects. In the case of visible light PDT, $^1O_2$ is considered an important reactive species for PDT because $^1O_2$ can be easily generated by visible light [41-44]. Critical targets of the generated $^1O_2$ include mitochondria and enzyme proteins. Moreover, DNA is also an important target biomolecule of photosensitized reactions [45-49]. Relevantly, photocatalytic $^1O_2$ generation by TiO_2 has been reported [38-40].

TiO_2, a nontoxic material, is chemically stable, and demonstrates a phototoxic effect. Therefore, an application of TiO_2 for PDT has been investigated [2]. The cytotoxicity of an illuminated TiO_2 film electrode for HeLa cells [18,19] and T-24 human bladder cancer cells [21] has been reported. Animal experiments also demonstrated the antitumor effect of TiO_2 NPs [20]. This report showed an antineoplastic effect on skin cancer in mouse models.

2. Photocatalytic DNA damage by TiO_2 NPs

Cellular DNA damage photocatalyzed by TiO_2 NPs was demonstrated by the experiment using cancer cells [18,19,21]. TiO_2 NPs can be taken into the cancer cell [27]; however, incorporation into the cell nucleus is difficult [18]. Therefore, it is speculated that the indirect mechanism contributes to DNA damage induced by photo-irradiated TiO_2 NPs. Hence, model experiments using isolated DNA were performed [31, 32]. In this section, an example of photocatalytic DNA damage by TiO_2 NPs was introduced.

2.1. Isolated DNA damage photocatalyzed by TiO_2 NPs and its sequence specificity

Photo-irradiated TiO_2 NPs catalyze DNA damage in the presence of copper(II) ion [31]. Relevantly, copper-aided photosterilization of microbial cells on TiO_2 was reported [8]. DNA damage by anatase NPs is more severe than that by rutile NPs. The DNA damage is enhanced by piperidine treatment, because photo-irradiated TiO_2 NPs cause not only DNA strand breakage but also base oxidation. In general, hot piperidine cleaves DNA strand at modified base. Photo-irradiated TiO_2 NPs induce the formation of piperidine-labile products at the bolded site of 5′-**T**G, 5′-T**G**, and 5′-**T**C (Figure 4). Furthermore, TiO_2 NPs photocatalyze DNA strand cleavage at the bolded guanines of 5′-T**G** and 5′-T**C** in a DNA fragment treated with *E coli* formamidopyrimidine-DNA glycosylase (Fpg protein), which can catalyze the excision of piperidine-resistant 8-oxo-7,8-dihydro-2′deoxyguanine (8-oxo-G) [50,51]. The formation of 8-oxo-G was confirmed by an analysis with a high-performance liquid chromatography (Figure 5). In addition, Fpg protein can cleave the oxidized cytosine, such as 5-hydroxy cytosine [52]. These results suggest that photo-irradiated TiO_2 NPs induce 8-oxo-G formation adjacent to piperidine-labile thymine lesions. Such double-base lesions should be generated from one radical hit that leads through a secondary reaction to a tandem base damage at pyrimidine and adjacent residues [53-56]. Actually, it has been reported that H_2O_2 induces tandem mutations in human cells via vicinal or cross-linked base modification in the presence of copper(II) ion [57]. Since repairing of cluster DNA damage in living cells is difficult [58], such clustered base damage, including double-base lesions, appears to play an important role in the phototoxicity of TiO_2 NPs.

2.2. Mechanism of DNA damage photocatalyzed by TiO_2 NPs

Catalase, a well-known scavenger of H_2O_2, and bathocuproines, a copper(I) ion chelator, inhibit DNA damage photocatalyzed by TiO_2 NPs, whereas, typical $OH^{\cdot}$ scavenger cannot inhibit the DNA damage. These results suggest that H_2O_2 and copper(I) ion participate in DNA damage

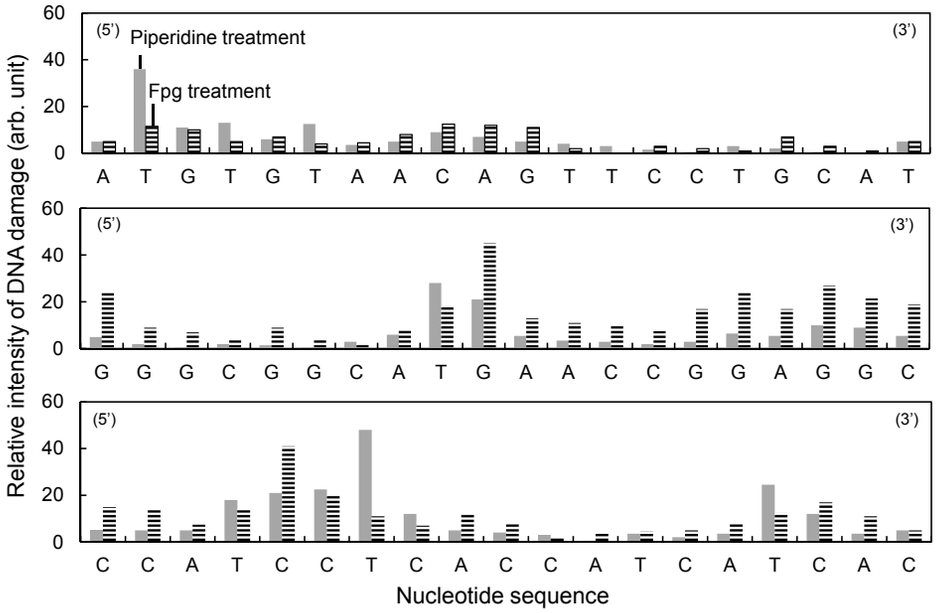

Figure 4. Sequence specificity of DNA damage photocatalyzed by anatase TiO_2 NPs. The ^{32}P-end-labeled 211 base pair DNA fragment (*p53* tumor suppressor gene) and 8 μg mL^{-1} anatase was irradiated with ultraviolet light (365 nm, 10 J cm^{-2}) with 20 μM copper(II) ion in a 10 mM sodium phosphate buffer (pH 7.8). After the photocatalytic reaction, the DNA fragments were treated with hot piperidine or Fpg and analyzed by an electrophoresis.

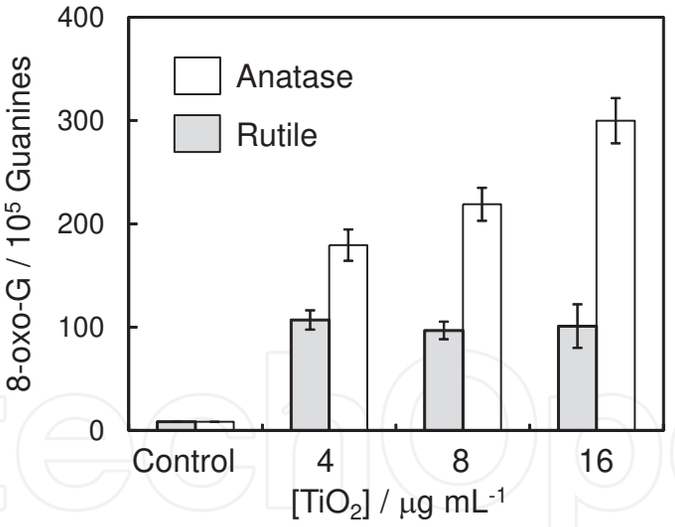

Figure 5. Formation of 8-oxo-G by the photocatalytic reaction of anatase or rutile NPs. Calf thymus DNA was treated by the photocatalytic reaction of anatase or rutile NPs (365 nm, 10 J cm^{-2}) with 20 μM copper(II) ion in a 10 mM sodium phosphate buffer (pH 7.8). After the photocatalytic reaction, the samples were analyzed with a high-performance liquid chromatography.

by photo-irradiated TiO_2 NPs. It has been reported that $OH^{\cdot}$ is not the main reactive species involved in DNA damage by H_2O_2 and copper(I) ions [34-36, 59]. DNA-associated copper(I) ions may generate other oxidants, including a copper–peroxo intermediate, such as Cu(I)-OOH, which is generated from the reaction of H_2O_2 and copper(I) ions [34-36, 59]. Indeed,

methional, which can scavenge Cu(I)-OOH [36, 59], shows inhibitory effect on DNA damage photocatalyzed by TiO_2 NPs. The generation of these reactive species may be responsible for the formation of piperidine-labile products and 8-oxo-G.

On the other hand, a high concentration of anatase NPs can catalyze DNA photodamage without copper(II) ions. Typical $OH^{\cdot}$ scavengers, ethanol and sugars, effectively inhibit the DNA photodamage by a high concentration of anatase NPs. The DNA damage induced by photo-irradiated anatase NPs without copper(II) ions is observed at every nucleobases without site specificity. Such DNA damage without sequence-specificity is the typical pattern of $OH^{\cdot}$-mediated DNA damage [34].

A proposed mechanism of DNA damage photocatalyzed by TiO_2 NPs is shown in Figure 6. The crystalline forms of TiO_2, anatase and rutile, are semiconductors with band gap energies of 3.26 and 3.06 eV, which correspond to the following wavelengths of light: 385 and 400 nm, respectively. When a TiO_2 semiconductor NPs absorbs photon with energy greater than their band gap, electrons in the valence band are excited to the conduction band, creating electron-h^+ pairs and causing various chemical reactions [2-5]. The electron acts as a reductant, whereas the h^+ is a powerful oxidant. In aqueous environments, oxygen molecule can be reduced by the electron into $O_2^{\cdot -}$, and water molecule can be oxidized by the h^+ into $OH^{\cdot}$. In general, formed $O_2^{\cdot -}$ can be dismutated into H_2O_2 by proton. The oxygen reduction may precede the reduction of copper(II) ions under aerobic condition, since the concentration of dissolved oxygen is higher (~250 μM) than that of the copper(II) ion used in this study (20 μM). The copper(II) reduction may be mediated by $O_2^{\cdot -}$. Hydrogen peroxide reacts with copper(I) ions to generate other oxidants, including a copper–peroxo intermediate, resulting in the oxidation of DNA bases. Copper ions, which are essential components of chromatin [60,61], are found to bind DNA with high affinity [62,63]. Therefore, copper ions may play an important role in reactive oxygen generation *in vivo*, although mammals have evolved means of minimizing the levels of free copper ions and most copper ions bind to protein caries and transporters [64]. Hydroxyl radicals formed by the reaction of water with an h^+ in the valence band of TiO_2 NPs also slightly participate in DNA damage photocatalyzed by anatase NPs. Because $OH^{\cdot}$ is a strong oxidative agent, $OH^{\cdot}$ can damage every nucleobase [34]. The present results suggest that H_2O_2 mainly participate in the phototoxicity of TiO_2 NPs and the contribution of $OH^{\cdot}$ is relatively small. Fujishima *et al.* also reported the involvement of H_2O_2 generated from $O_2^{\cdot -}$ in the cytotoxicity of illuminated TiO_2 NPs [2-4, 8-13].

TiO_2 NPs might be a potential agent for PDT [22-25]. TiO_2 NPs can be incorporated into cancer cells and demonstrate cytotoxicity under photo-irradiation [2-4, 26-28]. Photocatalytic reaction by TiO_2 NPs induces a number of functional changes in cell including altered permeability of cellular membranes to potassium and calcium ions, release of RNA and proteins and cytotoxicity [2,18-21]. It has been reported that DNA can be a target biomolecule of the photocatalytic reaction of TiO_2 NPs [26-30]. Although incorporation of TiO_2 NPs into cell nucleus is difficult [18], the generated H_2O_2 by a photocatalytic reaction of TiO_2 NPs can be easily diffused and incorporated in a cell nucleus, leading to DNA photodamage with metal ions. Relevantly, several studies demonstrated that DNA is a potential target of PDT [47,65,66]. Therefore, the

http://dx.doi.org/10.5772/61302

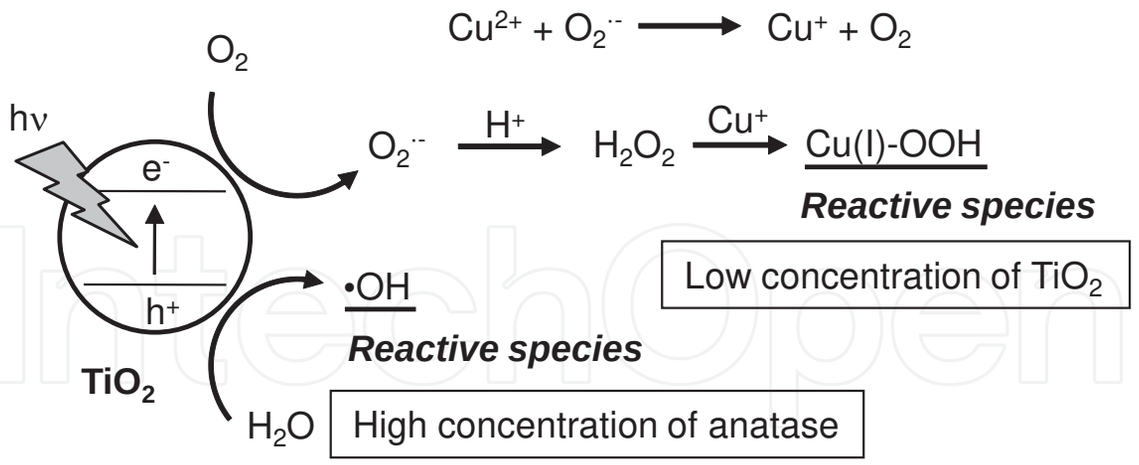

Figure 6. Proposed mechanism of DNA damage photocatalyzed by TiO_2 NPs.

metal-mediated DNA damage through the photocatalysis of TiO_2 NPs may participate in cytotoxicity by photo-irradiated TiO_2 NPs.

3. Secondary production of reactive oxygen species from photocatalyzed materials by TiO_2 NPs

As mentioned above, DNA damage in human cells by TiO_2 NPs has also been reported [26-28]. The direct DNA damage by TiO_2 NPs photocatalyst *in vitro* has been also studied [31, 32]. However, the DNA-damaging mechanism *in vivo* is not well-understood because the incorporation of the TiO_2 NPs in the cell nucleus is difficult [18]. Since the TiO_2 photocatalytic reaction occurs in a complex biological environment, an interaction between TiO_2 NPs and biomaterials may participate in the generation of reactive species to induce DNA damage. Hence, the effect of sugars, which are ubiquitous biomaterials, on DNA damage photocatalyzed by TiO_2 NPs was examined [37].

In the case of anatase, a high concentration of TiO_2 NPs can damage DNA at every nucleobase by $OH^{\cdot}$ generation in the absence of copper(II) ions. Typical free $OH^{\cdot}$ scavengers inhibited this copper(II)-independent DNA damage. These results indicate that free $OH^{\cdot}$ partly contributes to DNA damage photocatalyzed by TiO_2. On the other hand, scavengers of $OH^{\cdot}$, such as a sugar (mannitol), ethanol, and formate, enhanced the copper(II)-dependent DNA damage [31]. These scavengers themselves did not induce DNA damage. Since $OH^{\cdot}$ can oxidize most biomaterials, the oxidized products of biomaterials by the TiO_2 photocatalyst may damage DNA via the generation of secondary reactive oxygen species. The addition of sugars, glucose and galactose, which are ubiquitous biomolecules, enhanced the DNA damage photocatalyzed by TiO_2 NPs. Enhancement of DNA damage by sugars has seldom been reported, and these sugars themselves could not induce DNA damage. Therefore, the products of the photocatalytic reaction of these sugars by TiO_2 NPs is responsible for the copper(II)-dependent damage to DNA. Indeed, the glucose and galactose oxidized by the TiO_2 photocatalytic reaction caused

DNA damage in the presence of copper(II) ion [37]. The inhibitory effect of various scavengers for DNA damage by the photo-oxidized products of sugars by TiO_2 was examined. Catalase inhibited DNA damage by the photocatalyzed glucose, indicating the involvement of H_2O_2. Bathocuproine, which is a chelator of copper(I) ion, also inhibited DNA damage by the photocatalyzed glucose, suggesting the involvement of copper(I) ion. The free $OH^{\cdot}$ scavengers had no or little inhibitory effect on DNA damage. The inhibitory effect of superoxide dismutase (SOD) was weak, suggesting that $O_2^{\cdot -}$ itself is not the main reactive species for DNA damage. Similar results were observed in the case of galactose. Fluorometry using folic acid [67] demonstrated the formation of H_2O_2 from the photocatalyzed sugars (Figure 7). The amount of H_2O_2 generation was comparable with that of other H_2O_2-mediated DNA-damaging drugs [68]. H_2O_2 generation was not observed in the absence of copper(II) ions. These results showed that the oxidized products of sugars generate H_2O_2 during the reaction with copper(II) ions, resulting in secondary DNA damage.

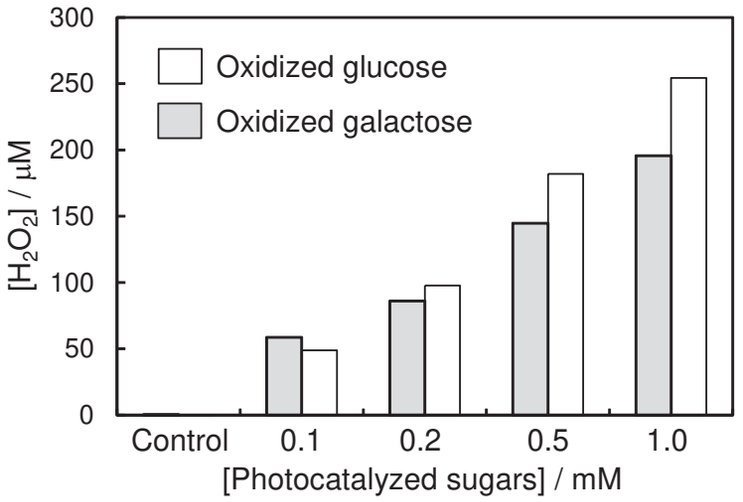

Figure 7. Hydrogen peroxide generation from photo-oxidized glucose and galactose by TiO_2 NPs. The buffer solution with 10 mM sugars was previously irradiated (365 nm, 6 J cm^{-2}) with 100 μg mL^{-1} anatase NPs. The TiO_2 NPs were removed by centrifugation, and the solution containing the oxidized sugars was used. One mL of solution containing the treated sugars and 10 μM of folic acid was incubated (60 min, 37 °C) in the presence of 20 μM copper(II) chloride, and the fluorescence intensity was measured (excitation: 360 nm, detection: 450 nm). The concentration of the generated H_2O_2 was determined by the calibration curve method.

These sugars act as an electron donor for the photocatalytic reaction [15,37]. Partially oxidized sugars, such as aldehyde compounds, are possibly produced through this photocatalytic oxidation. The mechanism of DNA damage by the photocatalyzed product of sugars is proposed in Figure 8. Aldehydes can generate H_2O_2 via its further oxidation [69], though these sugars themselves are stable compounds. Many studies have reported DNA damage by H_2O_2 and copper(II) ions [34-36, 70]. Various chemical compounds, including aldehydes, easily produce $O_2^{\cdot -}$ through their autoxidation process. The autoxidation is markedly enhanced by copper(II) ion, which is an essential component of chromatin [60, 61]. The formed $O_2^{\cdot -}$ is rapidly dismutated into H_2O_2. Although the generated H_2O_2 itself cannot damage DNA, H_2O_2 reduces copper(II) into copper(I), leading to the activation of H_2O_2 through the formation of reactive

species, such as Cu(I)-OOH [34-36, 59]. Indeed, methional, a scavenger of Cu(I)-OOH, inhibited the DNA damage. This reactive species cannot be scavenged by the free $OH^{\cdot}$ scavengers; however, it can effectively oxidize the nucleobases [34-36, 59].

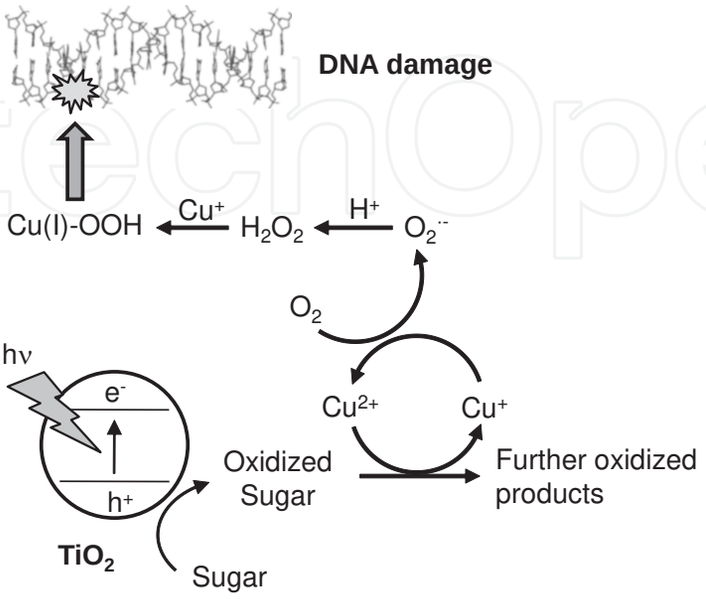

Figure 8. Proposed mechanism of secondary DNA damage by photocatalyzed sugars.

Although TiO_2 is not likely to be incorporated in a cell nucleus [18], H_2O_2 generated via a photocatalytic reaction can be easily diffused and incorporated in a cell nucleus. This DNA-damaging mechanism via H_2O_2 generation may participate in the phototoxicity of TiO_2. *In vivo*, the cell membrane is an important reaction field for the TiO_2 photocatalyst because TiO_2 NPs show affinity with a cell membrane [18]. Further, a part of the TiO_2 NPs can become incorporated into the cell. Sugars on the cell membrane and cytoplasm may be oxidized by the TiO_2 photocatalytic reaction. The generated h^+ and $OH^{\cdot}$ can oxidize these sugars, leading to the formation of secondary H_2O_2 from their photo-oxidized products.

In summary, sugars enhance the DNA damage photocatalyzed by TiO_2 NPs. This enhancement of DNA damage is due to the secondary generation of a reactive oxygen species, H_2O_2, which can diffuse in the cell and damage cellular DNA. These findings suggest that the secondary H_2O_2 generation contributes to the phototoxicity of TiO_2 more than the direct formation of reactive oxygen species does.

4. Singlet oxygen formation through photocatalytic reaction of TiO_2 NPs

A contribution of $^1O_2$ in the TiO_2 photocatalytic reaction was reported [38]. Singlet oxygen generation by TiO_2 photocatalysis has been demonstrated by the emission measurement of $^1O_2$, which is assigned to the transition from $^1O_2(^1\Delta_g)$ to $^3O_2(^3\sum_g)$ [39, 40]. Because $^1O_2$ is considered to be an important reactive species in PDT process [22-25], the clarification of the

contribution of $^1O_2$ generated by TiO_2 photocatalysis is closely related to a design of photocatalyst for medicinal application. Thus, $^1O_2$ generation in the TiO_2 photocatalysis and its importance on biomolecular damage was examined [40].

The typical emission of $^1O_2$ at around 1270 nm was observed during irradiation of TiO_2 NPs. Relatively strong emission of $^1O_2$ was observed in nonpolar organic solvents such as dichloromethane. The quantum yield (Φ_Δ) of $^1O_2$ generation by TiO_2 photocatalysis in ethanol was estimated from the comparison of $^1O_2$ emission intensities by TiO_2 NPs and methylene blue (Φ_Δ= 0.52) [71] and the absorbance of the TiO_2 NP dispersions. Because the scattering by suspended TiO_2 NPs makes the calculation of absorbed light intensity complex, the precise estimation of the Φ_Δ is difficult. Thus, the Φ_Δ was estimated using the apparent absorbance of TiO_2 NPs. The calculated value indicates the lowest limit of the Φ_Δ by TiO_2 photocatalysis in ethanol. The reported lifetime of $^1O_2$ generated via TiO_2 photocatalytic reaction is 5 μs [39]. This value is shorter than that by the photosensitized reaction of methylene blue (12 μs) [72]. Since the emission intensity of $^1O_2$ is proportional to its lifetime, the Φ_Δ was corrected by the lifetime of $^1O_2$. The estimated value of Φ_Δ by both types of TiO_2, anatase and rutile, was about 0.02 in ethanol. This value of Φ_Δ is enough large to induce oxidative damage to biomolecules. The $^1O_2$ emission in D_2O was completely quenched by the addition of SOD, which is the enzyme to dismutate $O_2^{\cdot-}$ into H_2O_2. These results can be explained by the fact that $^1O_2$ is formed by the reoxidation of $O_2^{\cdot-}$, generated from the photoreduction of oxygen molecules by TiO_2 NPs (Figure 3). The intensity of $^1O_2$ emission observed in the case of rutile was significantly larger than that by anatase in D_2O. The difference of the $^1O_2$ generation by these two types of TiO_2 crystalline forms can be reasonably explained by that in aqueous solution. H_2O_2 generation proceeds in the photocatalysis of anatase rather than $O_2^{\cdot-}$ generation, whereas $O_2^{\cdot-}$ is the main product from oxygen photoreduction mediated by rutile [17]. These results support the mechanism of $^1O_2$ generation via $O_2^{\cdot-}$ by TiO_2 photocatalysis.

The emission spectrum of $^1O_2$ by TiO_2 (in both, anatase and rutile type cases) slightly blue-shifted (~4 nm) compared with that by methylene blue. These results suggest that the surroundings of the $^1O_2$ generated on the TiO_2 surface are different from that by methylene blue in solution. In the case of the photosensitization of methylene blue, the generated $^1O_2$ deactivates in the homogeneous media of solvents. A possible explanation of the blue-shift is that most of the $^1O_2$ generated by TiO_2 NPs deactivates on the TiO_2 surface.

The intensity of $^1O_2$ emission by TiO_2 photocatalysis in liposome was significantly larger than that in an aqueous solution in both, anatase and rutile type cases. The enhancement of the $^1O_2$ emission can be explained by the elongation of the lifetime of $^1O_2$ or the acceleration of the photocatalytic reaction. This result shows that phospholipids membrane is an important environment of the phototoxic reaction mediated by $^1O_2$ in the photocatalytic reactions of TiO_2 NPs. Indeed, high affinity of TiO_2 NPs with a cell membrane was reported [18]. Consequently, an environmental effect of a cell membrane is important for the photocatalytic reaction of TiO_2 NPs. Since amino acid residues in proteins can be oxidized by $^1O_2$ [42], a membrane protein should be the target biomolecule in cell membrane. Indeed, $^1O_2$ emission was quenched by the addition of bovine serum albumin, a typical water soluble protein, suggesting scavenging of the $^1O_2$ generated by TiO_2 photocatalysis through oxidation of protein.

In vivo, nicotinamide adenine dinucleotide (NADH) is one of the most important target biomolecule oxidized by $^1O_2$ [73, 74]. NADH demonstrates the typical absorption peak at around 340 nm in an ultraviolet absorption spectral measurement, and this absorption band is diminished by the oxidation. It has been reported that TiO_2 NPs hardly induce the oxidation of NADH in aqueous solution during ultraviolet irradiation. Since NADH hardly adsorbed on a surface of TiO_2 NPs, the $^1O_2$ could not effectively oxidize NADH in solution. As mentioned above, it has been reported that photo-irradiated TiO_2 NPs can induce DNA damage mainly through H_2O_2 and $OH^·$, and the $^1O_2$-mediated DNA damage could not be observed [31]. These reports concluded that the photocatalytic $^1O_2$ generation on the surface of TiO_2 NPs is not important in the damaging mechanism of the biomolecules such as DNA and NADH, of which the affinity with TiO_2 surface is small.

In conclusion, photo-irradiated TiO_2 NPs can produce $^1O_2$ through reoxidation of $O_2^{·-}$, which is formed by photocatalytic reduction of oxygen molecule on the surface of TiO_2 NPs. Since most of the $^1O_2$ deactivated on TiO_2 surface, the $^1O_2$ on TiO_2 surface cannot induce the oxidation of DNA and NADH. However, the $^1O_2$ generation by TiO_2 photocatalysis could be enhanced in the microenvironment of phospholipids membrane. These findings suggest that $^1O_2$ may contribute to phototoxicity of TiO_2 NPs through oxidation of membrane protein.

5. Conclusions

TiO_2 NPs photocatalyze DNA oxidation. A relatively small concentration of TiO_2 NPs frequently induces tandem base oxidation at guanine and thymine residues through H_2O_2 generation in the presence of a copper(II) ion. A copper–peroxo complex is considered to be an important reactive species responsible for this DNA damage. In addition, cytosine residues are also photooxidized by TiO_2 NPs. In the case of a high concentration of TiO_2 NPs, $OH^·$ contributes to DNA damage without sequence specificity. In the presence of sugars, TiO_2 NPs indirectly induce DNA damage by the secondary H_2O_2, which is produced through an autoxidation process of the photo-oxidized products of sugars by TiO_2 NPs. Furthermore, $^1O_2$ is also produced by photo-irradiated TiO_2 NPs. The $^1O_2$ generation is explained by the reoxidation of $O_2^{·-}$, which is produced by photocatalytic reduction of the oxygen molecule adsorbed on the surface of TiO_2 NPs. The photocatalyzed formation of $^1O_2$ might contribute to the oxidation of the membrane protein. These mechanisms of photocatalytic reactive oxygen formation should be involved in the photocytotoxicity of TiO_2 NPs. Because TiO_2 is a chemically stable and nontoxic material, the bactericidal activity and cytotoxicity against cancer cells will play more important roles in the field of medical applications of nanomaterials.

Acknowledgements

The author wishes to thank Professor Shosuke Kawanishi (Suzuka University of Medical Science) for his helpful discussion about DNA damage. The reported works were supported

by a Grant-in-Aid for Scientific Research on Priority Areas (417) from the Ministry of Education, Culture, Sports, Science, and Technology (MEXT) of the Japanese Government.

Author details

Kazutaka Hirakawa[1,2]

Address all correspondence to: hirakawa.kazutaka@shizuoka.ac.jp

1 Applied Chemistry and Biochemical Engineering Course, Department of Engineering, Graduate School of Integrated Science and Technology, Shizuoka University, Johoku, Naka-ku, Hamamatsu, Shizuoka, Japan

2 Department of Optoelectronics and Nanostructure Science, Graduate School of Science and Technology, Shizuoka University, Johoku, Naka-ku, Hamamatsu, Shizuoka, Japan

References

[1] Fujishima A, Honda K. Electrochemical photolysis of water at a semiconductor electrode. *Nature*. 1972;238:37-38. DOI: 10.1038/238037a0

[2] Fujishima A, Rao TN, Tryk DA. Titanium dioxide photocatalysis. *J Photochem Photobiol C: Photochem Rev*. 2000;1:1-21. DOI: 10.1016/S1389-5567(00)00002-2

[3] Fujishima A, Zhang X, Tryk DA. TiO_2 photocatalysis and related surface phenomena. *Surf Sci Rep*. 2008;63:515-582. DOI: 10.1016/j.surfrep.2008.10.001

[4] Liu K, Cao M, Fujishima A, Jiang, J. Bio-inspired titanium dioxide materials with special wettability and their applications. *Chem Rev*. 2014;114:10044-10094. DOI: 10.1021/cr4006796

[5] Tachikawa T, Fujitsuka M, Majima T. Mechanistic insight into the TiO_2 photocatalytic reactions: design of new photocatalysts. *J Physic Chem C*. 2007;111:5259-5275. DOI: 10.1021/jp069005u

[6] Kikuchi Y, Sunada K, Iyoda T, Hashimoto K, Fujishima A. Photocatalytic bactericidal effect of TiO_2 thin films: dynamic view of the active oxygen species responsible for the effect. *J Photochem Photobiol A*. 1997;106:51-56. DOI:10.1016/S1010-6030(97)00038-5

[7] Sunada K, Kikuchi Y, Hashimoto K, Fujishima A. Bactericidal and detoxification effects of TiO_2 thin film photocatalysts. *Environ Sci Technol*. 1998;32:726-728. DOI: 10.1021/es970860o

[8] Sato T, Taya M. Copper-aided photosterilization of microbial cells on TiO_2 film under irradiation from a white light fluorescent lamp. *Biochem Engin J*. 2006;30: 199-204. DOI: 10.1016/j.bej.2006.04.002

[9] Page K, Palgrave RG, Parkin IP, Wilson M, Savin SLP, Chadwick AV. Titania and silver-titania composite films on glass-potent antimicrobial coatings. *J Mater Chem*. 2007;17:95-104. DOI: 10.1039/B611740F

[10] Page K, Wilson M, Parkin IP. Antimicrobial surfaces and their potential in reducing the role of the inanimate environment in the incidence of hospital-acquired infections. *J Mater Chem*. 2009;19:3819-3831. DOI: 10.1039/B818698G

[11] Dunnill CW, Aiken ZA, Kafizas A, Pratten J, Wilson M, Morgan DJ, Parkin IP. White light induced photocatalytic activity of sulfur-doped TiO_2 thin films and their potential for antibacterial application. *J Mater Chem*. 2009;19:8747-8754. DOI: 10.1039/B913793A

[12] Mansfield CM, Alloy MM, Hamilton J, Verbeck GF, Newton K, Klaine SJ, Roberts AP. Photo-induced toxicity of titanium dioxide nanoparticles to *Daphnia magna* under natural sunlight. *Chemosphere*. 2014;120:206-210. DOI:10.1016/j.chemosphere.2014.06.075

[13] Ljubas D, Smoljanić G, Juretić H. Degradation of Methyl Orange and Congo Red dyes by using TiO2 nanoparticles activated by the solar and the solar-like radiation. *J Environ Manage*. 2015;161:83-91. DOI: 10.1016/j.jenvman.2015.06.042

[14] St. John MR, Furgala AJ, Sammells AF. Hydrogen generation by photocatalytic oxidation of glucose by platinized n-titania powder. *J Physic Chem*. 1983;87:801-805. DOI: 10.1021/j100228a021

[15] Enea O. Structural and pH effects on the photo- oxidation of some polyalcohols and sugars by the suspensions of TiO_2 particles. *Electrochim Acta*. 1986;31:405. DOI: 10.1016/0013-4686(86)80098-6

[16] Ishibashi K, Fujishima A. Quantum yields of active oxidative species formed on TiO_2 photocatalyst. *J Photochem Photobiol A*. 2000;134:139-142. DOI: 10.1016/S1010-6030(00)00264-1

[17] Goto H, Hanada Y, Ohno T, Matsumura M. Quantitative analysis of superoxide ion and hydrogen peroxide produced from molecular oxygen on photo-irradiated TiO_2 particles. *J Catalysis*. 2004;225:223-229. DOI: 10.1016/j.jcat.2004.04.001

[18] Cai R, Hashimoto K, Itoh K, Kubota Y, Fujishima A. Photokilling of malignant cells with ultra-fine TiO_2 powder. *Bull Chem Soc Jap*. 1991;64:1268-1273. DOI: org/10.1246/bcsj.64.1268

[19] Cai R, Hashimoto K, Kubota Y, Fujishima A. Increment of photocatalytic killing of cancer cells using TiO_2 with the aid of superoxide dismutase. *Chem Lett*. 1992:427-430. DOI: org/10.1246/cl.1992.427

[20] Cai R, Kubota Y, Shuin T, Sakai H, Hashimoto K, Fujishima A. Induction of cytotoxicity by photoexcited TiO_2 particles. *Canc Res*. 1992;52:2346-2348. DOI: org/content/52/8/2346

[21] Kubota Y, Shuin T, Kawasaki C, Hosaka M, Kitamura H, Cai R, Sakai H, Hashimoto K, Fujishima A. Photokilling of T-24 human bladder cancer cells with titanium dioxide. *Brit J Canc*. 1994;70:1107-1111. PMCID: PMC2033697

[22] Dolmans DEJGJ, Fukumura D, Jain RK. Photodynamic therapy for cancer. *Natur Rev Canc*. 2003;3:380-387. DOI: 10.1038/nrc1071

[23] Castano AP, Mroz P, Hamblin MR. Photodynamic therapy and anti-tumour immunity. *Natur Rev Canc*. 2006;6:535-545. DOI: 10.1038/nrc1894

[24] Wilson BC, Patterson MS. The physics, biophysics and technology of photodynamic therapy. *Phys Med Biol*. 2008;53:R61-R109. DOI: 10.1088/0031-9155/53/9/R01

[25] Collins HA, Khurana M, Moriyama EH, Mariampillai A, Dahlstedt E, Balaz M, Kuimova MK, Drobizhev M, Yang VXD, Phillips D, Rebane A, Wilson BC, Anderson HL. Blood-vessel closure using photosensitizers engineered for two-photon excitation. *Natur Photonics*. 2008;2:420-424. DOI: 10.1038/nphoton.2008.100

[26] Dunford R, Salinaro A, Cai L, Serpone N, Horikoshi S, Hidaka H, Knowland J. Chemical oxidation and DNA damage catalysed by inorganic sunscreen ingredients. *FEBS Lett*. 1997;418:87-90. DOI: 10.1016/S0014-5793(97)01356-2

[27] Wamer WG, Yin Jj, Wei RR. Oxidative damage to nucleic acids photosensitized by titanium dioxide. *Free Rad Biol Med*. 1997;23:851-858. DOI: 10.1016/S0891-5849(97)00068-3

[28] Meena R, Rani M, Pal P, Rajamani P. Nano-TiO_2-induced apoptosis by oxidative stress-mediated DNA damage and activation of p53 in human embryonic kidney cells. *Appl Biochem Biotechnol*. 2012;167:791-808. DOI 10.1007/s12010-012-9699-3

[29] Nakagawa Y, Wakuri S, Sakamoto K, Tanaka N. The photogenotoxicity of titanium dioxide particles. *Mutat Res*. 1997;394:125-132. DOI: 10.1016/S1383-5718(97)00126-5

[30] Kashige N1, Kakita Y, Nakashima Y, Miake F, Watanabe K. Mechanism of the photocatalytic inactivation of Lactobacillus casei phage PL-1 by titania thin film. *Curr Microbiol*. 2001;42:184-189. DOI: 10.1007/s002840010201

[31] Hirakawa K, Mori M, Yoshida M, Oikawa S, Kawanishi S. Photo-irradiated titanium dioxide catalyzes site specific DNA damage via generation of hydrogen peroxide. *Free Rad Res*. 2004;38:439-447. DOI:10.1080/1071576042000206487

[32] Tachikawa T, Asanoi Y, Kawai K, Tojo S, Sugimoto A, Fujitsuka M, Majima T. Photocatalytic cleavage of single TiO_2/DNA nanoconjugates. *Chem Eur J*. 2008;14:1492-1498. DOI: 10.1002/chem.200701030

[33] Neyens E, Baeyens J. A review of classic Fenton's peroxidation as an advanced oxidation technique. *J Hazardous Mater*. 2003;98:33-50. DOI: 10.1016/S0304-3894(02)00282-0

[34] Kawanishi S, Hiraku Y, Oikawa S. Mechanism of guanine-specific DNA damage by oxidative stress and its role in carcinogenesis and aging. *Mutat Res*. 2001;488:65-76. DOI: 10.1016/S1383-5742(00)00059-4

[35] Hiraku Y, Ito K, Hirakawa K, Kawanishi S. Photosensitized DNA damage and its protection via a novel mechanism. *Photochem Photobiol*. 2007;83:205-212. DOI: 10.1562/2006-03-09-IR-840

[36] Hirakawa K, Oikawa S, Hiraku Y, Hirosawa I, Kawanishi S. Catechol and hydroquinone have different redox properties responsible for their differential DNA-damaging ability. *Chem Res Toxicol*. 2002;15:76-82. DOI: 10.1021/tx010121s

[37] Hirakawa K. Titanium dioxide photocatalyzes DNA damage via the secondary generation of hydrogen peroxide in the presence of sugars. *Trends Photochem Photobiol*. 2012;14:69-73.

[38] Konaka R, Kasahara E, Dunlap WC, Yamamoto Y, Chien KC, Inoue M. Irradiation of titanium dioxide generates both singlet oxygen and superoxide anion. *Free Rad Biol Med*. 1999;27:294. DOI: 10.1016/S0891-5849(99)00050-7

[39] Nosaka Y, Daimon T, Nosaka AY, Murakami Y. Singlet oxygen formation in photocatalytic TiO_2 aqueous suspension. *Physic Chem Chemic Physic*. 2004;6:2917. DOI: 10.1039/B405084C

[40] Hirakawa K, Hirano T. Singlet oxygen generation photocatalyzed by TiO_2 particles and its contribution to biomolecule damage. *Chem Lett*. 2006;35:832-833. DOI: 10.1246/cl.2006.832

[41] Foote CS. Definition of Type I and Type II photosensitized oxidation. *Photochem Photobiol*. 1991;54: 659. DOI: 10.1111/j.1751-1097.1991.tb02071.x

[42] DeRosa MC, Crutchley RJ. Photosensitized singlet oxygen and its applications. *Coordination Chem Rev*. 2002;233-234:351-371. DOI: 10.1016/S0010-8545(02)00034-6

[43] Schweitzer C, Schmidt R. Physical mechanisms of generation and deactivation of singlet oxygen. *Chem Rev*. 2003;103:1685-1758. DOI: 10.1021/cr010371d

[44] Ogilby PR, Foote CS. Chemistry of singlet oxygen. 42. effect of solvent, solvent isotopic substitution, and temperature on the lifetime of singlet molecular oxygen ($^1\Delta_g$). *J Am Chem Soc*. 1983;105:3423-3430. DOI: 10.1021/ja00349a007

[45] Ravanat JL, Sauvaigo S, Caillat S, Martinez GR, Medeiros MH, Di Mascio P, Favier A, Cadet J. Singlet oxygen-mediated damage to cellular DNA determined by the comet assay associated with DNA repair enzymes. *Biol Chem*. 2004;385:17-20. DOI: 10.1515/BC.2004.003

[46] Cadet J, Ravanat JL, Martinez GR, Medeiros MH, Di Mascio P. Singlet oxygen oxidation of isolated and cellular DNA: product formation and mechanistic insights. *Photochem Photobiol*. 2006;82:219-225. DOI: 10.1562/2006-06-09-IR-914

[47] Hirakawa K. DNA damage through photo-induced electron transfer and photosensitized generation of reactive oxygen species. In: Kimura H, Suzuki A (eds.), *New Research on DNA Damage*. Nova Science Publishers Inc; 2008. p. 197-219. ISBN: 978-1-60456-581-2

[48] Hirakawa K, Hirano T, Nishimura Y, Arai T, Nosaka Y. Dynamics of singlet oxygen generation by DNA-binding photosensitizers. *J Physic Chem B*. 2012;116:3037-3044. DOI: 10.1021/jp300142e

[49] Hirakawa K, Nishimura Y, Arai T, Okazaki S. Singlet oxygen generating activity of an electron donor-connecting porphyrin photosensitizer can be controlled by DNA. *J Physic Chem B*. 2013;117:13490-13496. DOI: org/10.1021/jp4072444

[50] Kasai H, Crain PF, Kuchino Y, Nishimura S, Ootsuyama A, Tanooka H. Formation of 8-hydroxyguanine moiety in cellular DNA by agents producing oxygen radicals and evidence for its repair. *Carcinogenesis*. 1986;7:1849-1851. DOI: 10.1093/carcin/7.11.1849

[51] Ito K, Inoue S, Yamamoto K, Kawanishi S. 8-Hydroxydeoxyguanosine formation at the 5' site of 5'-GG-3' sequences in double-stranded DNA by UV radiation with riboflavin. *J Biologic Chem*. 1993;268:13221-13227.

[52] D'Ham C, Romieu A, Jaquinod M, Gasparutto D, Cadet J. Excision of 5,6-dihydroxy-5,6-dihydrothymine, 5,6-dihydrothymine, and 5-hydroxycytosine from defined sequence oligonucleotides by Escherichia coli endonuclease III and Fpg proteins: kinetic and mechanistic aspects. *Biochemistry*. 1999;38:3335-3344. DOI: 10.1021/bi981982b

[53] David-Cordonnier MH, Laval J, O'Neill P. Clustered DNA damage, influence on damage excision by XRS5 nuclear extracts and Escherichia coli Nth and Fpg proteins. *J Biologic Chem*. 2000;275:11865-1873. DOI: 10.1074/jbc.275.16.11865

[54] Bourdat AG, Douki T, Frelon S, Gasparutto D, Cadet J. Tandem base lesions are generated by hydroxyl radical within isolated DNA in aerated aqueous solution. *J Am Chemic Soc*. 2000;122:4549-4556. DOI: 10.1021/ja994282i

[55] Box HC, Budzinski EE, Dawidzik JB, Gobey JS, Freund HG. Free radical-induced tandem base damage in DNA oligomers. *Free Rad Biol Med*. 1997;23:1021-1030. DOI: 10.1016/S0891-5849(97)00166-4

[56] Frelon S, Douki T, Favier A, Cadet J. Hydroxyl radical is not the main reactive species involved in the degradation of DNA bases by copper in the presence of hydrogen peroxide. *Chem Res Toxicol*. 2003;16:191-197. DOI: 10.1021/tx025650q

[57] Lee DH, O'Connor TR, Pfeifer GP. Oxidative DNA damage induced by copper and hydrogen peroxide promotes CG→ TT tandem mutations at methylated CpG dinu-

cleotides in nucleotide excision repair-deficient cells. *Nucleic Acids Res*. 2002;30:3566-3573.

[58] Blaisdell JO, Wallace SS. Abortive base-excision repair of radiation-induced clustered DNA lesions in Escherichia coli. Proc *Natl Acad Sci USA*. 2001;98:7426-7430. DOI: 10.1073/pnas.131077798

[59] Yamashita N, Murata M, Inoue. S, Burkitt MJ, Milne L, Kawanishi S. Alphatocopherol induces oxidative damage to DNA in the presence of copper(II) ions. *Chem Res Toxicol*. 1998;11:855-862. DOI: 10.1021/tx970129v

[60] Dijkwel PA, Wenink PW. Structural integrity of the nuclear matrix: differential effects of thiol agents and metal chelators. *J Cell Sci*. 1986;84:53-67.

[61] Saucier MA, Wang X, Re RN, Brown J, Bryan SE. Effects of ionic strength on endogenous nuclease activity in chelated and nonchelated chromatin. *J Inorgan Biochem*. 1991;41:117-124. DOI: 10.1016/0162-0134(91)80005-3

[62] Theophanides T, Anastassopoulou J. Copper and carcinogenesis. *Crit Rev Oncol Hematol*. 2002;42:57-64. DOI: 10.1016/S1040-8428(02)00007-0

[63] Bar-Or D, Thomas GW, Rael LT, Lau EP, Winkler JV. Asp-Ala-His-Lys (DAHK) inhibits copper-induced oxidative DNA double strand breaks and telomere shortening. *Biochem Biophys Res Commun*. 2001;282:356-360. DOI: 10.1006/bbrc.2001.4533

[64] Linder MC. Copper and genomic stability in mammals. *Mutat Res*. 2001;475:141-152. DOI: 10.1016/S0027-5107(01)00076-8

[65] Akhlynina TV, Jans DA, Rosenkranz AA, Statsyuk NV, Balashova IY, Toth G, Pavo I, Rubin AB, Sobolev AS. Nuclear targeting of chlorin e6 enhances its photosensitizing activity. *J Biol Chem*. 1997;272:20328-20331. DOI: 10.1074/jbc.272.33.20328

[66] Bisland SK, Singh D, Gariépy J. Potentiation of chlorin e6 photodynamic activity in vitro with peptide-based intracellular vehicles. *Bioconjugate Chem*. 1999;10:982-992. DOI: 10.1021/bc990020u

[67] Hirakawa K. Fluorometry of hydrogen peroxide using oxidative decomposition of folic acid. *Anal Bioanal Chem*. 2006;386:244-248. DOI: 10.1007/s00216-006-0649-1

[68] Hirakawa K, Midorikawa K, Oikawa S, Kawanishi S. Carcinogenic semicarbazide induces sequence-specific DNA damage through the generation of reactive oxygen species and the derived organic radicals. *Mutat Res*. 2003;536:91-101. DOI: 10.1016/S1383-5718(03)00030-5

[69] Mizutani H, Oikawa S, Hiraku Y, Murata M, Kojima M, Kawanishi S. Distinct mechanisms of site-specific oxidative DNA damage by doxorubicin in the presence of copper(II) and NADPH-cytochrome P450 reductase. *Canc Sci*. 2003;94:686-691. DOI: 10.1111/j.1349-7006.2003.tb01503.x

[70] Kawanishi S, Hiraku Y, Murata M, Oikawa S. The role of metals in site-specific DNA damage with reference to carcinogenesis. *Free Rad Biol Med*. 2002;32:822-832. DOI: 10.1016/S0891-5849(02)00779-7

[71] Usui Y, Kamogawa K. A standard system to determine the quantum yield of singlet oxygen formation in aqueous solution. *Photochem Photobiol*. 1974;19:245-247. DOI: 10.1111/j.1751-1097.1974.tb06506.x

[72] Merkel PB, Nilsson R, Kearns DR. Deuterium effects on singlet oxygen lifetimes in solutions. new test of singlet oxygen reactions. *J Am Chem Soc*. 1972;94:1030-1031. DOI: 10.1021/ja00758a072

[73] Petrat, F, Pindiur S, Kirsch M, de Groot H. NAD(P)H, a primary target of $^1O_2$ in mitochondria of intact cells. *J Biologic Chem*. 2003:278:3298-3307. DOI: 10.1074/jbc.M204230200

[74] Petrat F, Pindiur S, Kirsch M, de Groot H. "Mitochondrial" photochemical drugs do not release toxic amounts of $^1O_2$ within the mitochondrial matrix space. Arch Biochem Biophysics. 2003;412:207-215. DOI: 10.1016/S0003-9861(03)00063-8